

Detection of Trichophyton benhamiae with Fluorescence In Situ Hybridization

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Infrodu	JCTION

The following is one of the **adjusted protocols**:



The objective of this work was to detect **T. benhamiae** in **FFPE sections** of skin explants from experimentally infected guinea pigs using Fluorescence In Situ Hybridization.

T. benhamiae is a **zoonotic reemerging pathogen** causing dermatophytosis in humans.

Particularly affected are children due to close contact to pets. The condition is characterized by **dermatitis** especially in the face with severe inflammatory changes and pruritus known as Tinea faciei^{1,2}.

The In situ Hybridization is a common detection method already used for other pathogenetic fungi^{3,4} including dermatophytes^{5,6} as well for FFPE sections 4,7,8,9. Therefore, it was necessary to modify and establish a

specific protocol for this dermatophyte.

Material and methods

Fluorescence In Situ Hybridization (abbr.: FISH) is a qualitative detection method using fluorescence-labeled DNA probes.

These **DNA** probes [Tab. 1] hybridize with the complementary target DNA in the tissue. The labeled signal is detected by a fluorescence microscope transforming the incoming signal into a visible picture. Due to the separately detected fluorescence channels, the results are **summation images** [Fig. 1].



Fig. 4: T. benhamiae is widely distributed in the cornified layers of the epidermis (probe: Oligo 2)

Unfortunately, the detected results were not reproducible. Especially the accessibility of target DNA in the FFPE slides seems to be a barrier for the probes. Although the DNA probes produced a correct signal when applied directly on T. benhamiae samples, the results for the FFPE slides were inconsistent. Instead of pepsin digestion we tried **proteinase K and LysingEnzyme** (Sigma). Furthermore, the duration of the heat treatment was raised up to 20 minutes as well as the hydrochloric acid treatment. Especially the heat treatment is a critical step, as it has a great impact on the hybridization success⁹. Another problem that occurred were DNA probes **bleeding through** every channel of the microscope or at

wrong locations even though negative controls were correct. Non-specific fluorescences were described in other cases⁴ as well. The hybridization buffer was replaced by one consisting out of formamide, SDS, dextran sulfate and salmon DNA to avoid mismatches.

Probe type	Name	Fluorescence Signal
Specific for T. benhamiae	Oligo 1 Oligo 2	green
Universal for fungi	Pan-fungal Z Pan-fungal Aman	green
Enzyme probes*	Subtilisin (3,6) Mcpa	green
Positive Control	Oligo 3 Oligo 4	red
Negative Control	Green Fluorescent Protein	green

Tab.1 : The different DNA probes

By means of a PCR amplification it was shown that T. benhamiae is expressing those enzymes.



Results and discussion

T. benhamiae infiltrates the **cornified layers** from the outside to the inside and also the hair follicles. The amount of distribution increases with the duration of infection.

The fluorescence signal of T. benhamiae depends on the used probes [Tab.1], the cell nuclei are shown in blue.





Fig. 3: probe: Oligo 1





Fig.5: Low fluorescence signal (probe: Pan-fungal Aman) Fig.6: Cloudy background (probe: Oligo 4)

Furthermore, there were slides with a weak fluorescence signal or/and a high background signal [Fig. 5, 6], a common problem for ISH with fungi¹⁰. The amount of washing steps was increased, it helped marginally.

Conclusion

FISH as a qualitative method can be transferred to detect T. benhamiae. The detection of T. benhamiae in guinea pig skin explants, e.g. skin sections thereof, resulted in non-

Fig. 1 : Separate images (A-C) of the fluorescence channels; summation image in D (probe: Oligo 3)

Fig. 2: probe: Subtilisin 6

reproducible stainings. Although several modifications to the protocol were made, the results stayed below expectations. As the current set-up seems vulnerable to minor modifications, an approach to solve the issue would be to design new probes.

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